

EFFECT OF PHYSICAL AND CHEMICAL MUTAGENS INDIVIDUAL AND IN COMBINATION IN M1 GENERATION OF COWPEA (VIGNA UNGUICULATA L.)

Rakhi Kumari^{1*}, A.K. Pal¹, K. Srivastava² and Sunil Singh³

^{1*}Department of Horticulture, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi, India.
²Department of Genetics and Plant Breeding, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi, India.

³Directorate of Plant Protection Quarantine & Storage, NH-IV, Faridabad-121001 (Haryana) India.

Abstract

Effect of Gamma rays, Ethyl Methane Sulphonate (EMS) and their combination treatments on seed germination, reduction in seedling height, pollen sterility, ovule sterility and plant survival till maturity were studied in the M_1 generation of two cowpea (*Vigna unguiculata L.*) cultivars. *viz.*, Kashi Kanchan and Kashi Unnati. Different doses / concentrations of Gamma rays (10 kR, 20 kR, 30 kR, 40 kR and 50 kR), EMS (0.01 M, 0.02 M, 0.03 M, 0.04 M and 0.05 M) and their combination (10 kR +0.02 M, 20 kR+0.02 M, 30 kR + 0.02 M, 40 kR + 0.02 M and 50 kR + 0.02 M) were used to analyze their effect on above mentioned traits in M_1 generation of both the genotypes. Data collected from M_1 generation showed minimum germination percentage and maximum reduction in seedling height were observed at higher doses of combined treatments in both the cultivars. Similarly, highest pollen and ovule sterility as well as lowest plant survival were also recorded at maximum combined dose of mutagens as compared to the other treatments in both the varieties. Deleterious effects were more pronounced in higher doses, indicating almost a linear relationship.

Key words: Mutagens, Gamma rays, Ethyl methane sulphonate (EMS), Cowpea, Kashi Kanchan and Kashi Unnati.

Introduction

Cowpea (Vigna unguiculata L.) is an essential food legume and an important component of cropping systems in the drier regions of the tropics covering parts of Asia and Oceania, the Middle East, Southern Europe, Africa, southern USA, and Central and South America (Singh et al., 2002). The seeds are a major source of dietary protein in most developing countries. Cowpea seeds are a major source of plant proteins and vitamins for man, feed for animals, and also a source of cash income. The young leaves and immature pods are eaten as vegetables. In India, cowpea is developed in a territory of 3.9 million hectares with a generation of 2.21 million tons. It is developed in the semi-parched areas of Rajasthan, Gujarat, Karnataka, Tamil Nadu and Maharashtra for the most part as grain legume. The mature cowpea seed contains 24.8 percent protein, 63.6 percent carbohydrate, 1.9 percent fat, 6.3 percent fibre, 0.00074 percent thiamine, 0.00042 percent Riboflavin and 0.00281 percent Niacin.

In order to improve yield and other polygenic characters, mutation breeding should be effectively utilized (Deepa lakshmi and Ananda kumar, 2004). Mutation induction has become an established tool in plant breeding to supplement existing germplasm and improve cultivars in certain specific traits (Kurobane *et al.*, 1979). Induced mutations represent the same kind of changes that occur from natural causes Govindan, (2000). Mutagenesis has been widely used as a potent method of enhancing variability for crop improvement (Singh and Singh, 2001). Induced mutation, using physical and chemical mutagen, is a way to generate genetic variation, resulting in the creation of new varieties with better characteristic (Wongpiyasatid, 2000).

Physical mutagens incorporate different sorts of radiation, *viz.*, "X-rays, gamma rays, alpha particles, beta particles, quick and warm (slow) neutrons and bright rays. Cobalt-60 is commonly utilized for the generation of Gamma rays. Gamma rays cause chromosomal and

quality mutations like X-rays." Gamma rays are the most energetic form of electromagnetic radiation; their energy level is from ten to several hundred kilo electron volts and they are considered as the most penetrating compared to other radiations (Kovacs *et al.*, 2002). The chemical mutagens can be divided into four groups: Alkylating Agents like Ethyl Methane Sulphonate (EMS), Base Analogues, Acridine Dyes, and Nitrous Acid and Hydroxylamine. They induce chemical modification of nucleotides by mispairing, base pair substitutions, small deletion and insertion in genomes. EMS induce a high rate of mutations in both micro and higher organisms (Freese, 1963) and sometimes the mutation frequencies exceed those obtained by radiation (Goud, 1967).

Materials and Methods

Dry, healthy and uniform sized seeds of cowpea cultivars *viz.*, Kashi Kanchan and Kashi Unnati were treated with gamma rays at 10, 20, 30, 40 and 50 kR doses, ethyl methane sulphonate at 0.01, 0.02, 0.03, 0.04 and 0.05 M concentrations and combination treatment at 10kR +0.02M, 20kR+0.02M, 30kR + 0.02M, 40kR + 0.02M and 50kR + 0.02M. One ninety five (195) seeds

in each set were presoaked for 6 h., in water initially. Then, the seeds were immersed for 6 h., in the requisite concentration of mutagen ethyl methane sulphonate. The treated seeds were washed thoroughly in running tap water to eliminate the residual effect of chemical. The gamma irradiated seed of both varieties with five different doses (10kR, 20kR, 30kR, 40kR and 50 kR) were presoaked in distilled water for 6 hrs and then treated with 0.02 M EMS for 6 hrs.

The treated seeds of gamma rays, ethyl methane sulphonate, combination treatments and control seeds were immediately sown in the field in a randomized block design (RBD) with three replications at proper spacing between row to row and plant to plant respectively. Remaining 15 seeds of each treatments including control were raised on moist blotter paper in the laboratory for observing data on seed germination, seedlings height reduction besides observation of pollen sterility, ovule sterility and plant Survival till Maturity in the field in M_1 generation.

Results and Discussion

In present investigation, combined mutagenic

Treatments	Germination Percentage		Seedling height reduction as percent of control		Pollen s Sterility (%)		Ovule Sterility (%)		Plant survival as percent of control	
		Kanchan	Unnati	Kanchan	Unnati	Kanchan	Unnati	Kanchan	Unnati	Kanchan
Control	95.00	94.00	0.00	0.00	0.00	0.00	0.00	0.00	100.0	100.0
10 kR Gamma rays	91.33	90.67	6.25	2.39	8.20	10.6	3.5	4.6	93.69	90.62
20 kR Gamma rays	89.00	88.67	8.68	5.16	17.6	21.4	9.3	10.4	88.05	86.35
30 kR Gamma rays	85.33	85.00	13.19	13.63	25.6	32.6	18.6	21.3	79.64	80.57
40 kR Gamma rays	81.00	80.67	16.67	24.13	32.8	40.6	24.9	27.9	61.56	73.17
50 kR Gamma rays	77.33	75.00	23.78	27.07	39.4	49.5	28.4	35.6	48.44	66.81
0.01 M EMS	93.00	92.00	3.13	3.68	5.8	9.8	9.3	3.9	95.68	96.46
0.02 M EMS	89.67	86.67	5.90	6.26	16.6	20.6	16.8	12.3	76.39	88.24
0.03 M EMS	79.33	75.00	10.07	20.07	24.4	36.4	21.0	24.8	67.46	80.05
0.04 M EMS	76.67	72.00	13.89	26.89	28.2	43.7	26.4	33.9	62.67	70.79
0.05 M EMS	72.00	70.33	20.49	32.78	33.0	46.3	30.9	40.6	59.17	63.44
$10 \mathrm{kR} + 0.02 \mathrm{M}$	91.00	90.00	4.69	4.60	19.7	22.4	10.2	13.2	91.68	84.82
(Gamma rays + EMS)										
20 kR + 0.02 M	81.33	86.33	6.94	14.55	27.6	27.6	19.5	24.9	83.89	75.57
(Gamma rays + EMS)										
$30 \mathrm{kR} + 0.02 \mathrm{M}$	78.00	76.00	11.11	22.65	36.3	40.2	24.3	30.6	73.36	57.62
(Gamma rays + EMS)										
$40 \mathrm{kR} + 0.02 \mathrm{M}$	72.67	74.00	17.53	25.78	42.6	47.7	27.9	38.4	60.43	52.98
(Gamma rays + EMS)										
50 kR + 0.02 M	66.00	60.67	23.96	34.44	49.5	51.4	32.4	45.6	57.72	50.81
(Gamma rays + EMS)										

Table 1: Effect of different doses / concentrations of Gamma rays and Ethyl Methane Sulfonate (EMS), individual and in combination treatments on growth characters of Kashi Kanchan and Kashi Unnati in M1 Generation.

treatment (gamma rays + EMS) had more drastic effect on percent seed germination in both the varieties than gamma rays and EMS treatments as compared to control (Table 1). This clearly shows that the mutagens have exerted an inhibitory effect on physiological and biological processes necessary for seed germination; they include enzyme activity on seed germination. Progressive decrease in seed germination by the mutagenic treatments has also been reported earlier by Uma and Salimath, (2001); Gaur et al., (2003); Nawale et al., (2006) in cowpea. Among the all mutagenic treatments, combination treatment, 50 kR + 0.02 M EMS caused the most adverse effects. Similar reduction was also obtained by Kumar et al., (2010) in cowpea, Ramya et al., (2014) in black gram, Kamble and Patil, (2014) in chickpea who observed that the combination of physical and chemical mutagens was drastic, in their effect. Point mutations, enzyme inhibitions and chromosomal aberrations occur due to EMS and gamma rays.

In the present study, the seedling height reduction as percent of control was increased with an increase in dose or concentration of mutagenic treatments. In both the genotypes the maximum reduction was observed in combination treatments than individual treatments. Similar increase in seedling injury with increased concentrations of mutagens has been reported by Khan, (1990) in Vigna mungo, Khan et al., (2006) in mungbean, Larik et al., (2009) in Sorghum, in cowpea. An inverse relationship between the doses / concentrations of mutagen and plant survival was observed in both the genotypes. Similarly as germination, the plant survival rate was drastically reduced with an increase in dose/concentration of all the mutagens. Mutagens reduced plant survival has also been reported by Blixt et al., 1963 and Dhanavel et al., 2008. Dose dependent reduction in plant survival was observed with gamma rays, EMS and combination of both mutagens, which is in conformity with the results of Nawale et al., (2006); Singh et al., (2013); Ramya et al., (2014); Nawale et al., (2006); Dhanavel et al., (2008). Reduction in plant survival after mutagenic treatments could be attributed to decrease in the assimilation mechanism (Quasteler and Baer, 1950), inhibition of mitosis and chromosomal damage (Guckel and Sparrow, 1961). This might have been due to the effect of mutagens on meristematic tissues of the seed. Morphological variations, especially leaf abnormalities are the indicators of effective mutagen treatment

Pollen sterility increased with increasing doses of gamma rays, EMS and combined treatments. These results are supported with those of the earlier researchers like Singh and Singh, (2014); Kumar and Verma, (2010)

and Kumar et al., (2010) who concluded that the combined treatment of gamma rays and EMS proved to be more effective in inducing abnormalities and pollen sterility as compared to individual treatment, on seeds of cowpea (Vigna unguiculata (L.) Walp.) in variety K 5269. Among the mutagenic treatments (50 kR gamma+ 0.02M EMS) produced maximum pollen sterility followed by gamma rays and EMS. Pollen sterility and ovule sterility increased with increasing dose / concentration of Gamma rays Ethyl methane sulphonate. The findings were in conformity to there by Lambat et al., (2012) in Chilli. According to Konzak et al., (1961), Sparrow and Woodweel, (1962) and Sudhakaran, (1971) induction of the pollen sterility is due to chromosomal irregularities introduced by the mutagen. Siddiq and Swaminathan, (1969) concluded that chromosomal aberrations particularly high frequency of translocations was responsible for high sterility. In M1 generation, it was expected, as the chemical compounds are known to cause high pollen sterility (Blixt et al., 1963; Ramya et al., 2014). The dose of mutagen and ovule sterility showed inverse relationship with increasing doses of mutagens. The maximum ovule sterility was observed at highest dose or concentration of the mutagens. The findings were in conformity to there by Vanniarajan et al., (1993); Ahmed john, (1996) and Sagade and Apparao, (2012) in blackgram; Singh and Singh, (2007) in green gram.

Conclusion

All the growth traits were affected by different doses / concentrations of gamma rays and ethyl methane sulphonate either singly or in combination. Minimum germination percentage and maximum reduction in seedling height were observed at higher doses of combined treatments (50 kR + 0.02 M EMS) in both the cultivars. Similarly, highest pollen and ovule sterility as well as lowest plant survival were also recorded at a maximum combined dose of 50 kR + 0.02 M EMS as compared to the other treatments in both the varieties. Gamma rays belong to ionizing radiation and interact with atoms or molecules to produce free radicals in cells. These radicals can damage or modify important components of plant cells and have been reported to affect differentially the morphology, anatomy, biochemistry and physiology of plants depending on the irradiation level. Chemical mutagens usually cause point mutation, but the loss of a chromosome segment or deletion can also occur.

References

Ahmed John, S. (1996). Induced pollen and seed fertility in blackgram. *Madras Agricultural Journal.*, 83(5): 320-321.
Blixt, S. (1963). Studies on induced mutation in pea, VIII.

Ethylene imine and Gamma rays treatments of the variety Witham Wonder, *Agri. Hort. Genet.*, **22**:177-183.

- Deepalakshmi, A.J. and C. Anandakumar (2004). Creation of genetic variability for different polygenic traits in black gram (*Vigna mungo* (L.) Hepper) through induced mutagenesis. *Legume Res.*, 27(3): 188-192.
- Gaur, L.B., S.P. Singh and K. Srivastava (2013). Frequency and spectrum of chlorophyll mutation in chilli (*Capsicum annuum* L.). *International. J. of Plant Sci.*, **8**(1): 70-74.
- Govindan, A. (2000). Studies on induction of mutations on black grams (*Vigna mungo* (L.) varieties and hybrid populations and mutational effect of genetic parameters through generation. Ph.D., Thesis. India. Indian Institute of Pulses Research, Kanpur Project Co-ordinate's Report (2011). AICRP on MULLARP, Indian Institute of Pulses Research, Kanpur.
- Goud, J.V. (1967). Induced mutation in bread wheat, *Ind. J. Genet.*, **27**:40-55.
- Gunckel, J.E. and A.H. Sparrow (1961). Ionising radiations. Biochemical, physiological and morphological aspects of their effects on plants. *Ency. Pl. Physiol.*, 16: 365-387.
- Jayamani, P., N. Kumaravadivel, N. Nadarajan, A.R. Muthiah, C. Durairaj, A. Kamalakannan, S. Pazhanivelan and K. Thiyagarajan (2012). TNAU Blackgram CO6: A High Yielding Short Duration Variety. *Madras Agric. J.*, 99(1-3): 34-36.
- Jayanthi, S. (1986). Biological effects of gamma rays and Ethyl Methane Sulphonate in the M1 generation of Redgram (*Cajanus Cajan* (L.)) M.Sc.,(Agric) Thesis, Kerala University, India, J. Cytol. Genet., 6: 155-160.
- Kovacs, E. and A. Keresztes (2002). Effect of gamma and UV-B/ C radiation on plant cell. *Micron.*, **33**: 199-210.
- Kamble, G.C. and A.S. Patil (2014). Comparative mutagenicity of EMS and gamma radiation in wild chickpea. *International J. Sci., Environ. Tech.*, 3(1): 166-180.
- Khan, S. (1990). Studies on chemical mutagenesis in mungbean (*Vigna radiata* (L.) Wilczek). Ph.D. Thesis. Aligarh Muslim University, Aligarh.
- Khan, S. and M.R. Wani (2005). Genetic variability and correlation studies in chickpea mutants.
- Khan, S. and M.R. Wani (2006). MMS and SA induced genetic variability for quantitative traits in mungbean; *Indian J. Pulses Res.*, **19**(1): 50-52.
- Konzal, C.F., R.A. Nilan, R.R. Legault and R.E. Heiner (1961). Modification of induced genetic damage in seeds, In: Symp. on effects of ionizing radiation on seeds, IAEA, Karlsruhe, 155 -169.
- Kumar, G. and S. Verma (2010). Individual and combined mutagenesis of EMS and gamma rays in *Vigna unguiculata L. J. Plant Sci. Res.*, **26(2):** 159-163.
- Kurobane, I., H. Yamaguchi, C. Sander and R. Nilan (1979). The effects of gamma irradiation on the production and secretion of enzymes and enzymatic activities in barley. *Env. Exp. Botany.*, **19:** 75-84.
- Larik, A.S., S. Memon and Z.A. Soomro (2009). Radiation induced polygenic mutations in *Sorghum bicolor* L. J.

Agric. Res., 47(1): 11-19.

- Lgnacimuthu, S. and C.R. Babu (1989). Induced chromosomal abnormality and pollen sterility in wild and cultivated urd and mung beans. *Cytologio.*, **51**(1):159-167.
- Mathusamy, A. and N. Jayabalan (2002). Effect of mutagens on pollen fertility of cotton (*Gossypium hirsutum* (L)). *Indian J. Genet.*, **62(21):** 187.
- Mensah, J.K. and P.A. Akomeah (1992). Mutagenetic effects of hydroxylamine and streptomycin on the growth and yield of Cowpea (*Vigna unguiculata* (L.)) Walp.) *Legume Res.*, **15**: 39-44.
- Nawale, S.R., U.B. Apte and B.B. Jadhav (2006). Effect of gammarays and ethyl methane sulfonate on seed germination and survival of seedlings in cowpea (*Vigna unguiculata* (L.) Walp). *J. of Arid Legumes.*, 3(1): 102-105.
- Quastler, H. and M. Baer (1950). Inhibition of plant growth by radiation. III. Successive radiation effects, homologous response. *BioL Abstracts.*, **24:** 30984.
- Ramya, B., G. Nallathambi and S. Ganesh Ram (2014). The effect of mutagens on M1 population of black gram (*Vigno mungo* L. Hepper). *African J. of Biotechnology.*, **13(8)**: 951-956.
- Sagade, A.B. and B.J. Apparao (2011). M1 generation studies in Urd bean (*Vigna mungo* (L.) Hepper). Asiam J. Exp. Biol. Sci., 2(2): 372-375.
- Siddiq, E.A. and M.S. Swaminathan (1969). Enhanced mutation induction and recovery caused by nitrosoguanidine in *Oryza sativa. Indian J. Genet. PI. Breed.*, 28(3): 297-300.
- Singh, M. and V.P. Singh (2001). Genetic analysis of certain mutant lines of urdbean for yield and quality traits in M4 generation. *Ind. J. Pulses Res.*, **14(1):** 60-62.
- Singh, J.V. and R.N. Arora (2002). Genetics and Breeding. In: Kumar, D., and Singh, N. B. (eds.) *Guar in India*. Jodhpur: *Indian Arid Legume Society.*, 817233-310-2.
- Singh, A.K. and A.K. Singh (2007). Biological influence of gamma rays and Ethyl Methane Sulphonate and their synergistic effects in mungbean. *Journal of Food Legumes.*, 20(2): 153-155.
- Singh, Kuldeep and M.N. Singh (2014). Effectiveness and efficiency of Gamma rays and Ethyl Methane Sulphonate (EMS) in mungbean. *J. Food Legumes.*, **26:** 25-28.
- Sparrow, A.K. and G.M. Woodwell (1962). Production of the sensitivity of the plants to chronic gamma radiation. *Rad. Bot.*, **2:** 9-26.
- Sudhakaran, I.V. (1971). Meiotic abnormalities induced by gamma rays in *Vinca rosea* L., *Cytologia.*, **36:** 67-79.
- Uma, M.S. and P.M. Salimath (2001). Effect of ionizing radiations on germination and emergence of cowpea seeds. *Karnataka J. Agri. Sci.*, **14(4):** 1063-1064.
- Vanniarajan, C., P. Vivekanandan and J. Ramalingam (1993). Gamma ray and ethyl methane sulphonate induced sterility in blackgram. *Crop Research.*, **6(3):** 560-562.
- Wongpiyasatid, A., S. Chotechuen and P. Hormchan (2000). Induced mutations in mungbean breeding Regional yield trail of mungbean mutant lines. *Kasetsart J. (Nat. Sci.).*, 34: 443-449.